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SYNTHESIS AND BIOLOGICAL ACTIVITY OF NOVEL TRICYCLIC β-LACTAMS

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Abstract: The synthesis and the preliminary data on microbiological activity of some tricyclic β -lactam derivatives (4), analogues of synthetic "trinems", is described. Compound 14, key intermediate in the synthesis of 18, was stereoselectively obtained from its unsubstituted analogue, 12b, in 4 steps through enolphosphate 13.

β-Lactams are agents of choice in the current therapeutical approach to most of the infectious diseases induced by bacteria both for their high efficacy and their extremely safe toxicological profile. However the rapid emergence of bacterial strains resistant to the most generally used members of this class of compounds has stimulated, over the years, research for novel β-lactam structures that could overcome the problem of resistance.

Tricyclic β -lactams, trinems 1 (formerly tribactams) first described by our group,² are new, totally synthetic antibacterial agents, stable to clinically isolated β -lactamases, whose potency and breadth of spectrum of action have been demonstrated both *in vitro* and *in vivo*. In particular, 2a (GV104326) showed a particularly good biological profile³ and, together with its metabolically labile ester 2b (GV118819) has been selected for development studies.

Fig. 1

As a part of an ongoing programme aimed at studying how structure activity relationships, among trinems, are influenced by the size of ring C, we became interested in the synthesis of the 5-membered ring derivatives of general structures 3 and 4. We have already reported our results on the synthesis of 4-thia-1-azatricyclo[6.2.0.0^{3,7}]dec-2-enes (3).⁴ Although the preliminary data on both chemical stability and biological activity on this class of compounds were not very encouraging, we decided to undertake the synthesis of the carbocyclic analogues with general structure 4.

The commercially available 4-acetoxyazetidinone 5⁵ (Scheme 1) was protected at the nitrogen with a TMS derivative then successfully reacted with 1.5 equivalents of 1-trimethylsilyloxy-1-cyclopentene to give, after removal of the nitrogen protecting group, a 2:1 mixture of epimers 7a and 7b in 75% overall yield.

Separation of isomers by flash chromatography was not possible at this stage but could be achieved after transformation of the mixture of 7a and 7b to the corresponding phosphoranes 8a and 8b according to the protocol established by Woodward. Phosphorane 8a was transformed into fully protected trinem 9a in moderate

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yield by heating it in toluene at 100 °C. The silyl protecting group was then removed with TBAF and acetic acid in THF⁷ and the allyl group with sodium 2-ethylhexanoate in the presence of catalytic amounts of Pd(PPh₃)₄.⁸ The crude sodium salt 11a could be purified by reverse phase preparative HPLC (Lichrosorb C-18, $10\mu m$, $20 \times 250mm$, water as eluant).

Scheme 1

TBS= t-BuMe₂Si, All= CH₂CH=CH₂
a. (CH₃)₃SiCl, (C₂H₅)₃N, THF, 0°C; b. (CH₃)₃SiOTf, 0°C, CH₂Cl₂, c. KF, CH₃OH, 75% overall from 5; d. OHCCOOAll, toluene, ref.; e. SOCl₂, lutidine, THF, -20 °C then PPh₃, lutidine, THF; f. toluene, 100°C; g. TBAF, AcOH, THF, 25% from 9a; g. Pd(PPh₃)₄, Na 2-ethylhexanoate

When phosphorane **8b** was heated under similar conditions applied for **8a**, **9b** could be isolated in low yield (21%). Particular care was taken in the isolation of the product. As compound **9b** is unstable on silica gel, its isolation necessitated buffering the silica gel with AcOEt:TEA 95:5 before the purification of the reaction mixture. Compound **9b** undergoes spontaneous decomposition on standing at -30 °C and extensive degradation occurred during our attempts to remove the silyl protecting group.

Previous experience with the 6-membered ring C (1) homologues led us to hypothesise that the introduction of a substituent at position C-4 of 4 should lead to the best compromise in terms of chemical and enzymatic stability, antibacterial potency, breadth of spectrum and *in vivo* properties. Bearing these considerations in mind we directed our efforts towards the synthesis of 4-methoxy trinem 4 with (45,75) absolute configuration at the corresponding stereogenic centres.

The mixture of isomers 7a and 7b (Scheme 2) was protected at the amide nitrogen with *t*-butyldimethylsilylchloride and TEA. At this stage, the two epimers could be separated by MPLC. Isomer 12b was then converted into the enolphosphate 13 by treatment of the lithium enolate of 12b with diethylphosphorochloridate⁹ at -78°C followed by removal of the silyl protecting group on the nitrogen then transformed into ketoazetidinone 14 according to a procedure established in our laboratories. Oxidation with 3-chloroperoxybenzoic acid followed by treatment of the crude reaction mixture after work-up with methanol gave ketoazetidinone 14. It is worth mentioning that the presence of the isomer of 14 with inverted configuration at the newly formed stereogenic centre could not be detected by NMR analysis of the reaction mixture after work-

up. The conversion of 14 to the final compound 18 was accomplished following conventional methods; compounds 16, 17 and 18 were found chemically stable and could be stored for months at -30°C.¹¹

Scheme 2

a. TBSCl, TEA, DMF, flash chromatography; b. LiN(TMS) $_2$ (1.5 eq), THF, -78°C then CIPO(OEt) $_2$ (1.5 eq); c. KF, MeOH, 0°C, 49% from 12b; d. m-CPBA, CH $_2$ Cl $_2$; e. MeOH, lutidine, 51% from 13; f. OHCCOOAll, toluene, ref.; g. SOCl $_2$, lutidine, THF; h. PPh $_3$, lutidine, THF, 47%; i. toluene, Δ , 58%; j. TBAF, AcOH, THF, 39%; k. Pd(PPh $_3$) $_4$, Na 2-ethylhexanoate, 43% (after prep. HPLC).

Table 1. In vitro antibacterial activity* (MIC μg/ml) of trinems 11a and 18 in comparison with Imipenem.

	S.a. 663	S.a. 853	S.a. 1131	E.faecalis 850	E.coli 1850	P.a. 1911	C.p. 615
Imipenem	0.06	0.12	4	2	0.5	4	0.03
11a	1	0.5	8	16	2	>32	1
18	0.25	0.5	8	2	0.5	32	0.12

* Minimum Inhibitory Concentrations (MIC) determined in Mueller Hinton broth: Anaerobes Schadler broth Inoculum . 5x10⁵ CFU/ml S.a. 663= Staphylococcus aureus 663; S.a. 853= Staphylococcus aureus 853 β-lactamases producing strains; S.a. 1131= Staphylococcus aureus 1131 methicillin-resistant E.faecalis 850= Enterobacter faecalis 850E; E.coli 1850= Escherichia coli 1850E; P.a. 1911= Pseudomonas aeruginosa 1911; C. p. 615 = Clostridium perfringens 615E.

As expected, compounds 11a and 18 were sufficiently stable and could be tested. The antibacterial activity of trinem 18 was similar to imipenem except against *Pseudomonas aeruginosa*. Compound 18 (as most of the 4-substituted trinems synthesized) was found more stable than imipenem and meropenem to human renal DHP-I. The initial microbiological results suggest that the trinems with general structure 4 represent an extremely promising class of antibacterial agents.

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- and purified by flash chromatography to give compound 14 as a colourless oil (630mg, 51%).

 11. All the compounds synthesized have been characterised by routine analytical techniques. Compound 9b: HNMR 300MHz(δ, ppm, CDCl₃): 5.95 (m, 1H), 5.40 (m, 1H), 5.23 (m, 1H); 4.75 (m, 2H), 4.17 (m, 1H + 1H), 3.39 (m, 1H), 3.24 (dd, 1H, J₁=7.5Hz, J₂=2.7Hz), 2.82 (m, 1H), 2.40-2.20 (m, 2H), 2.20-1.90 (m, 2H), 1.90-1.70 (m, 2H), 1.22 (d, 3H, J=6.3Hz), 0.88 (s, 9H), 0.08 (s, 6H). Compound 10a: IR (ν_{max}, cm⁻¹, nujol): 1759-1730 (broad, C=O), 1591 (C=C).

 1H-NMR 300MHz (δ, ppm, D₂O): 4.03 (m, 1H), 3.82 (dd, 1H, J₁=2.1Hz, J₂=8.1Hz), 3.40-3.28 (m, 1H), 3.16 (dd, 1H, J₁=2.1Hz, J₂=6.6Hz), 2.38-2.28 (m, 2H), 2.05-1.80 (m, 3H), 1.24 (m, 1H), 1.13 (d, 3H, J=6.3Hz). Compound 18: IR (ν_{max}, cm⁻¹, nujol): 1755 (C=O), 1587 (C=C).

 1H-NMR 300MHz (δ, ppm, D₂O): 4.70 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-10 (m, 1H), 4.13 (dd, 1H, J₁=10Hz, J₂=3.5Hz), 4.11 (m, 1H), 3.40-

3.30 (m, 2H), 3.14 (s, 3H), 2.34-2.20 (m, 1H), 1.85-1.50 (m, 2H), 1.20 (m, 1H), 1.11 (d, 3H, J=6.6Hz).